

Low Template DNA

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Statistical methods in forensic genetics 7-10 October 2013, Copenhagen

(slides adapted from Peter Gill)

(Advanced DNA Interpretation workshop ISFG 2013, Melbourne)

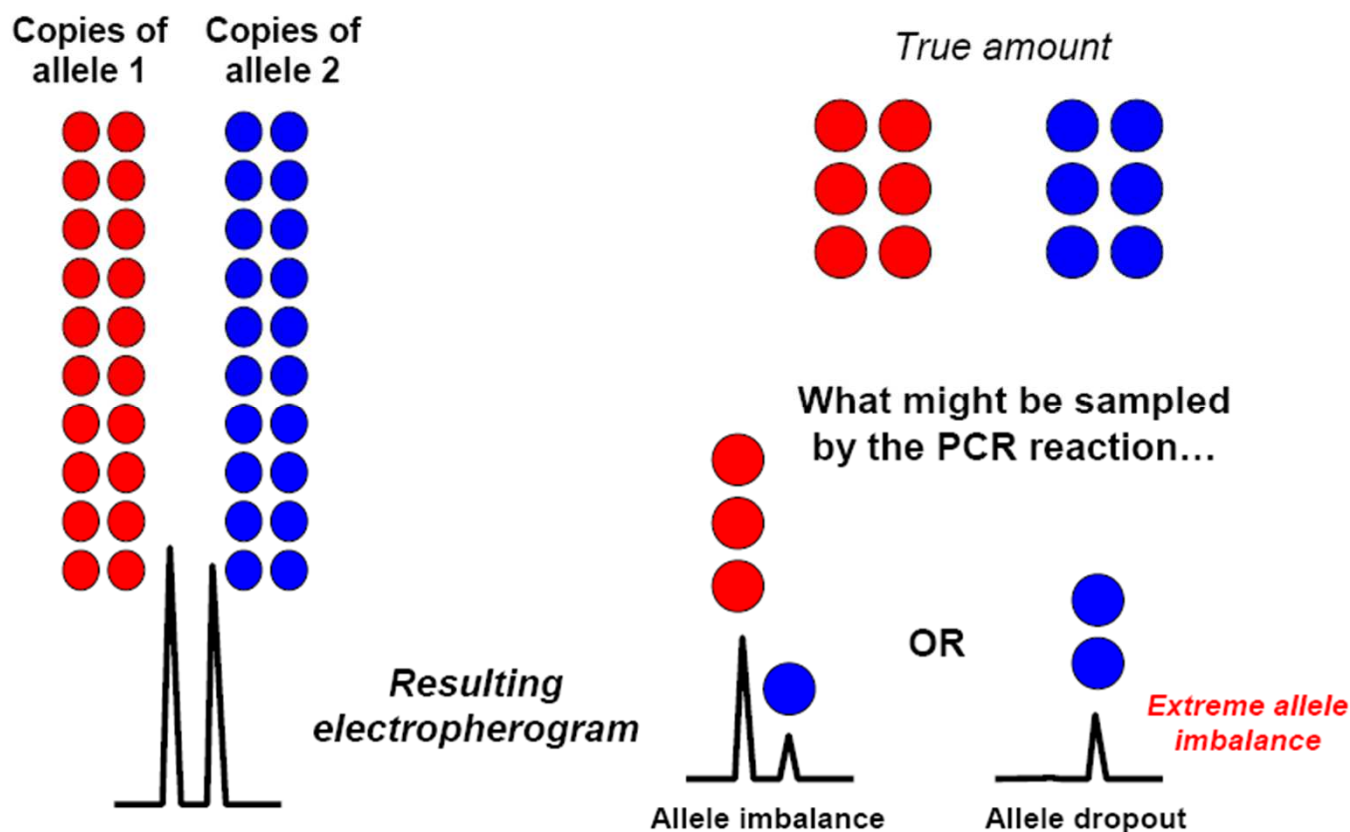
What is Low Copy Number?

- Let's make a list of what LCN is not
 - Its not related to an overall quantity of DNA (such as 200pg)
 - Its not restricted to 'touch DNA'
 - Its not related to any particular technique
- NY court found it to be a simple extension of an existing technique
- R. v. Reed accepted that the 34 cycle definition was not relevant to any definition of LT-DNA
- Why can't a definition be adduced?

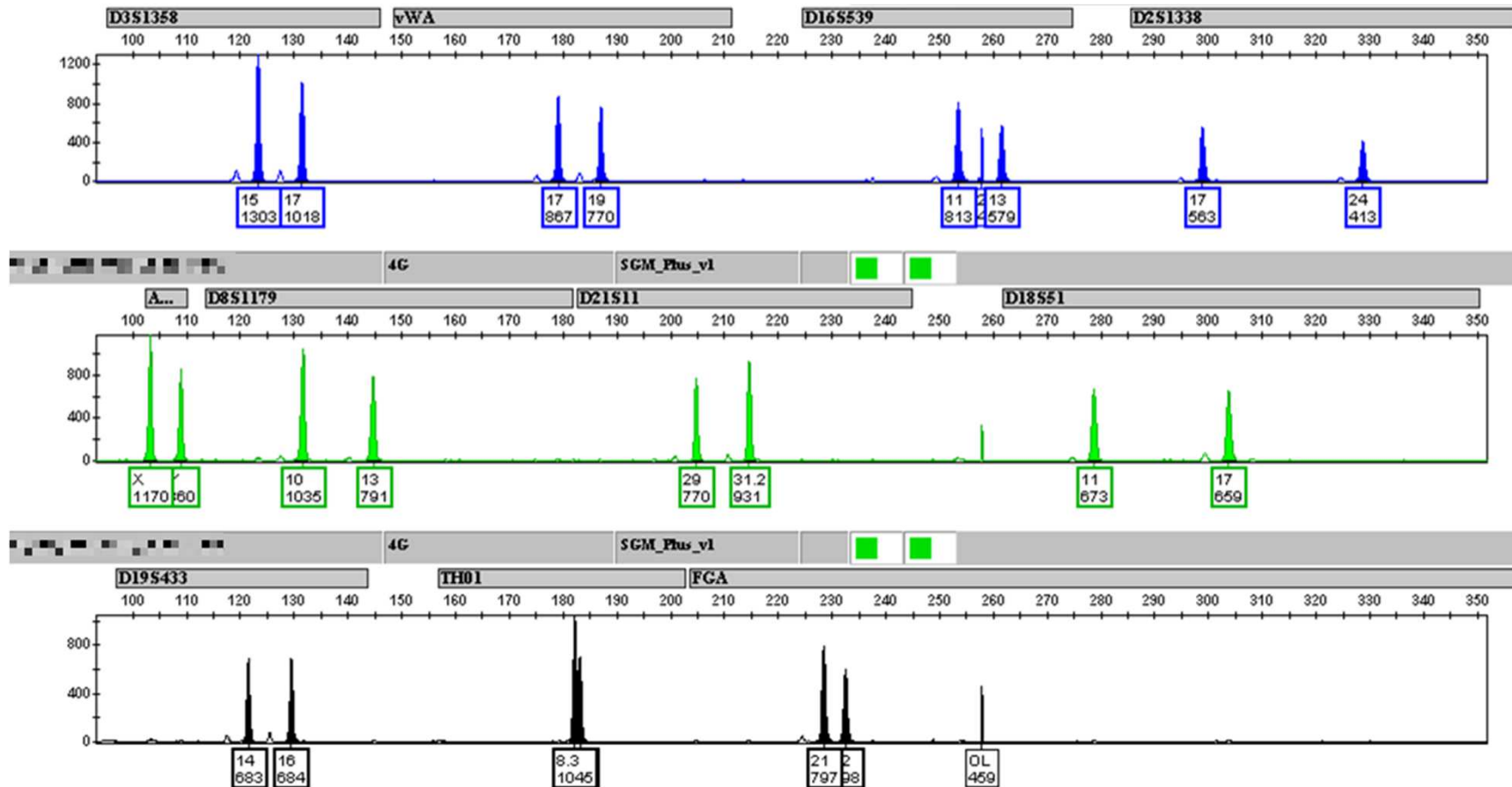
Stochastic variation

- In a heterozygous sample, one allele is amplified more than the other
- Leads to heterozygous imbalance or allele drop-out
 - Good quality DNA will always give heterozygous balance >60% i.e. both target alleles are amplified with similar efficiency
- Much more pronounced with low level DNA as there is less template DNA
- If one target gets amplified more in the first rounds of PCR then it becomes preferentially amplified in later rounds

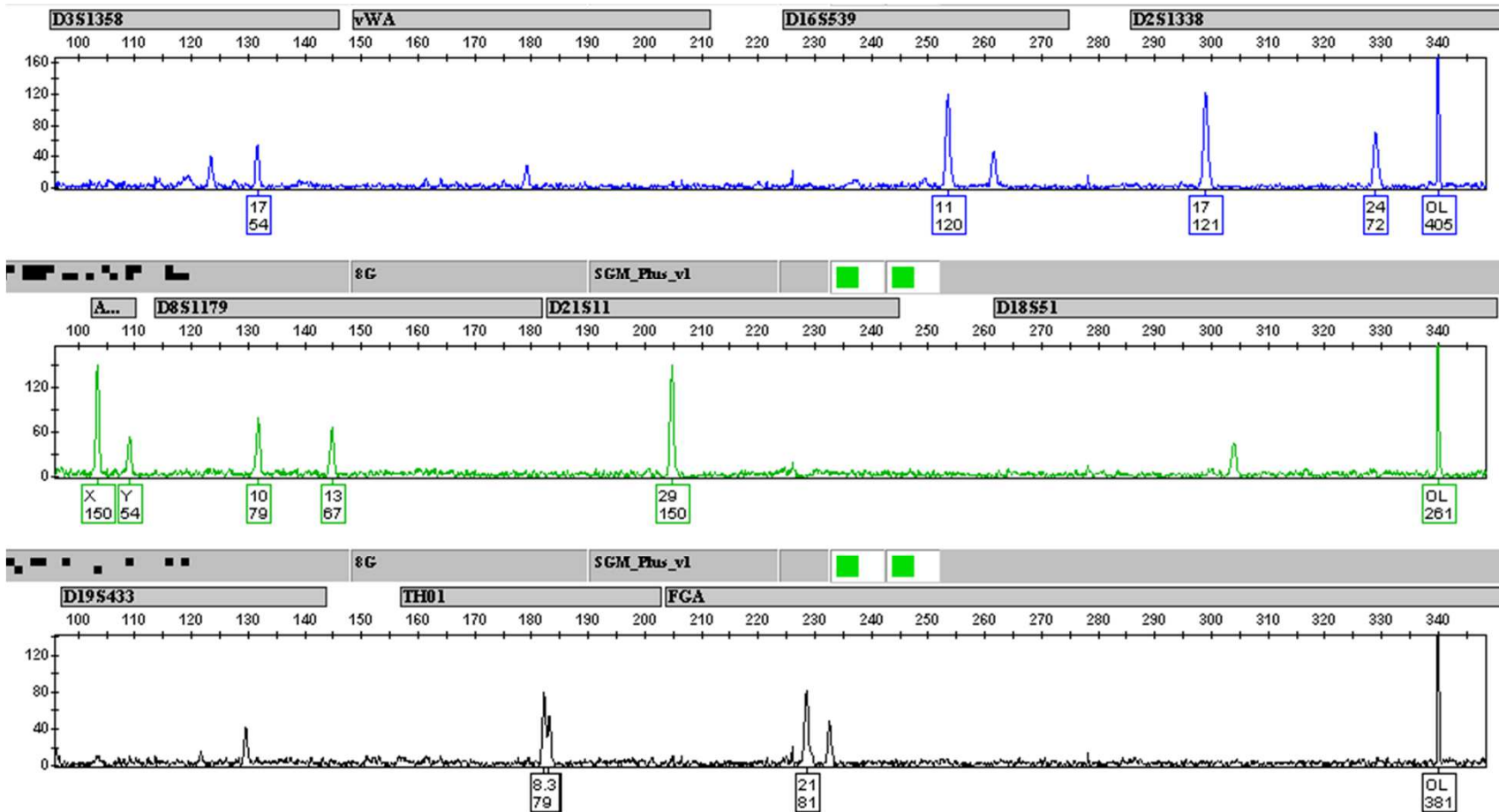
Heterozygous imbalance OR allele drop-out (from J Butler, NIST)



Example of good level DNA



Example of low level DNA



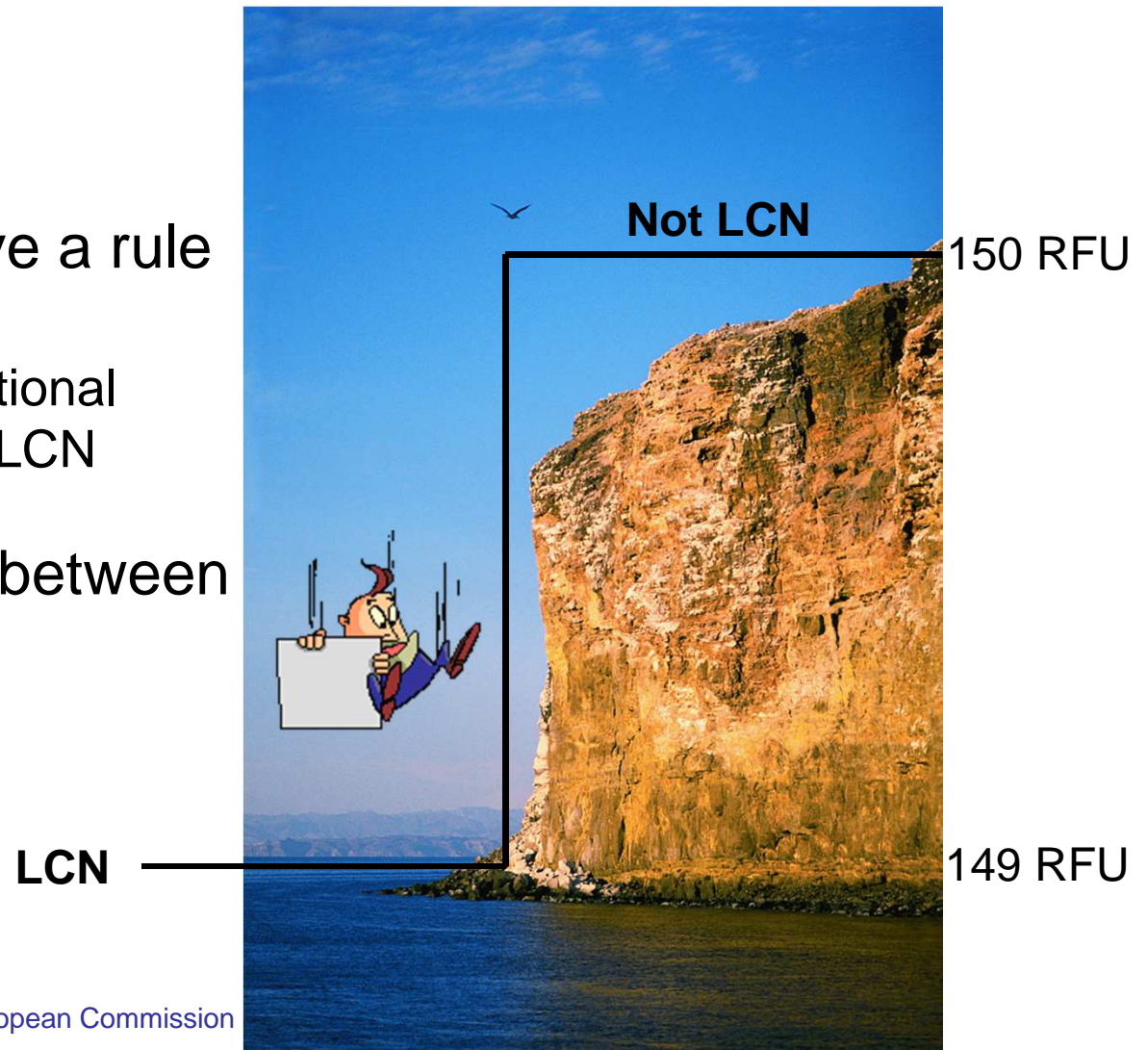
Illogical use of thresholds

- Falling off the cliff

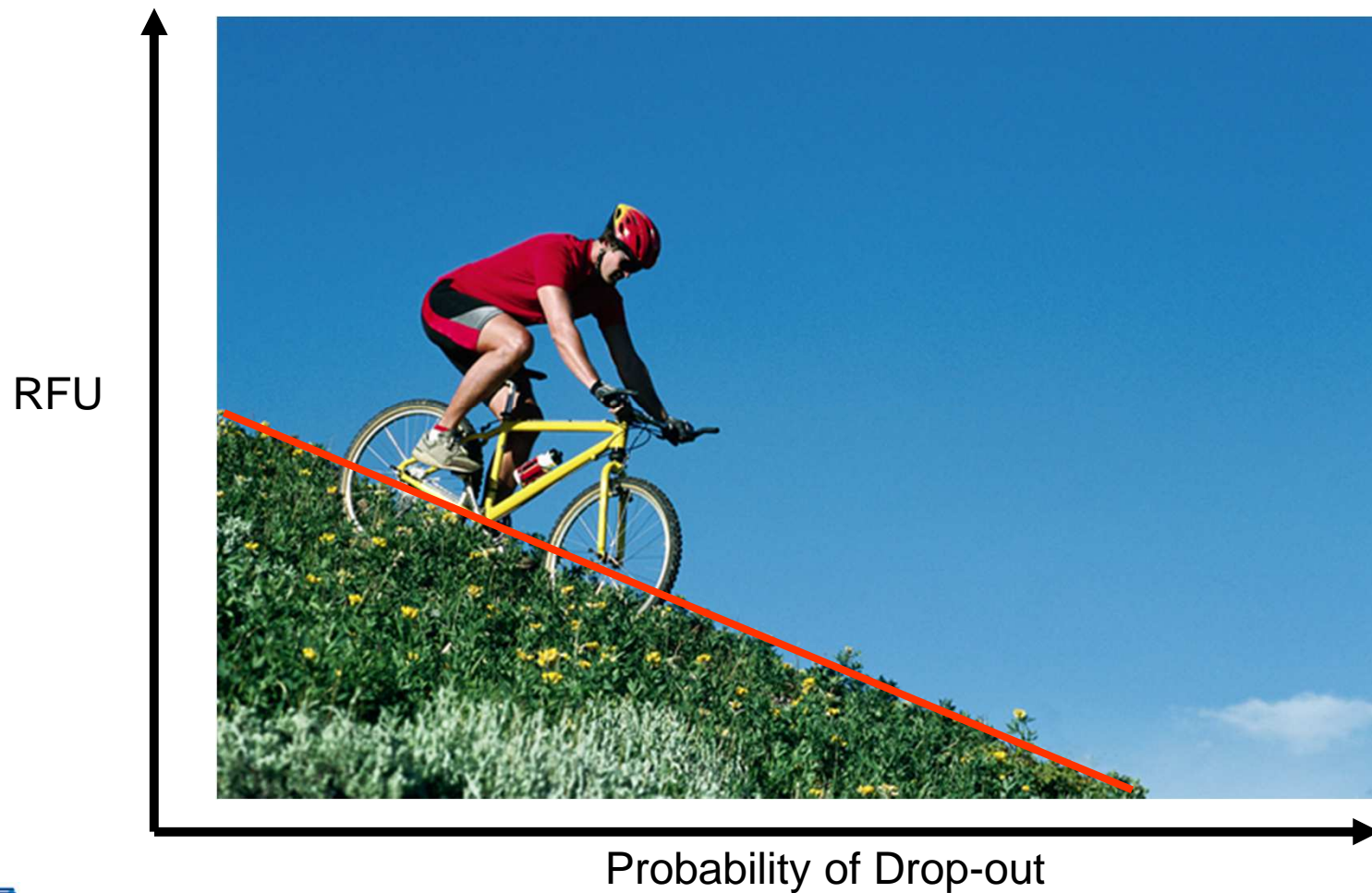
For example if we have a rule that states:

150 RFU – This is conventional
versus 149 RFU – This is LCN

- There is nothing in between



In reality it's a gentle ride downhill



Continuum of change

- The peak height rules break down when the quantity of DNA becomes very low – in particular the Hb guideline will no longer hold true
- Allele drop-out can lead to a heterozygous locus being genotyped as a homozygous locus
 - In standard DNA profiling, a homozygous peak height of 150 RFU is often used (stochastic threshold) i.e. single peaks <150 RFU are labeled 'F' indicating allele drop-out may have occurred

This is why we prefer a universal method

Forensic Science International: Genetics 4 (2010) 221–227



Contents lists available at ScienceDirect

Forensic Science International: Genetics

journal homepage: www.elsevier.com/locate/fsig



A universal strategy to interpret DNA profiles that does not require a definition of *low-copy-number*

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Papers outlining heterozygous balance

Holt CL, Buoncristiani M, Wallin JM, Nguyen T, Lazaruk KD, Walsh PS.
(2002) TWGDAM validation of AmpFISTR PCR amplification kits for forensic DNA casework. *J. Forensic Sci.* 47(1): 66-96.

Collins PJ, Hennessy LK, Leibel CS, Roby RK, Reeder DJ, Foxall PA.
(2004) Developmental validation of a single-tube amplification of the 13 CODIS STR loci, D2S1338, D19S433, and amelogenin: the AmpFISTR Identifiler PCR amplification kit. *J. Forensic Sci.* 49(6): 1265-1277.

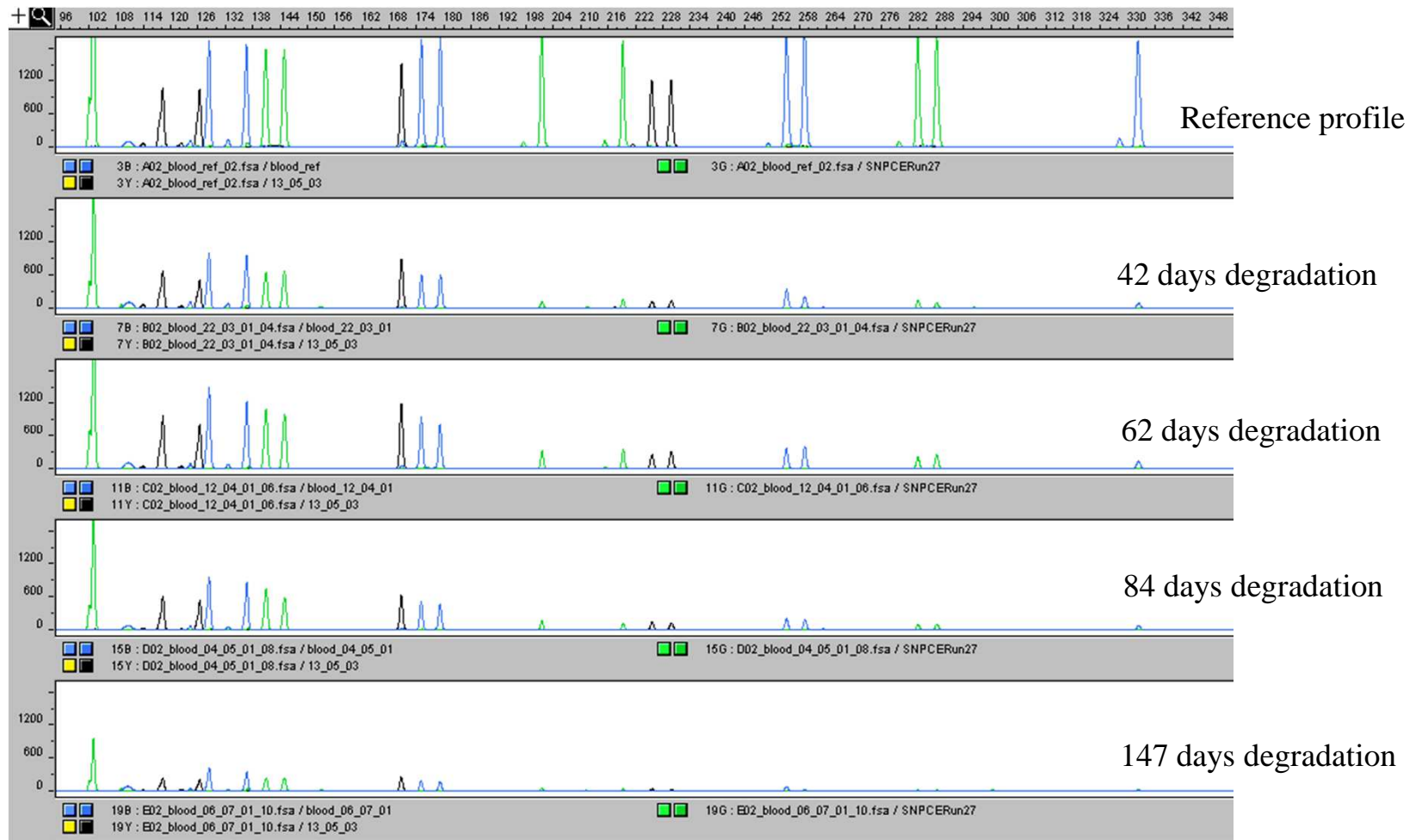
L.A. Dixon, C.M. Murray, E.J. Archer, A.E. Dobbins, P. Koumi & P. Gill
(2005) Validation of a 21-locus autosomal SNP multiplex for forensic identification purposes. *For. Sci. Int.* 154 (1): 62-77

Gill, P., Sparkes, R. and Kimpton, C. (1997). "Development of guidelines to designate alleles using an STR multiplex system." *Forensic Sci Int* **89**(3): 185-197

Degradation

- Occurs with fragmented / degraded DNA as there are more of the small target molecules available for amplification
- Leads to a distinctive slope in peak heights across the profile

Effect of degradation



Allele drop-in

- A contamination event resulting in only one or two foreign alleles
- Independent from gross contamination in that it comes from different sources

Contamination

- Gross contamination is identified as being from a single contributing source
- Dependent on transfer event as to when contamination occurred
- Could be pre-incident or post-incident

New methods

- Incorporate probability of dropout and dropin
- Uses statistical theory that is well established
- The theory can be used to evaluate complex mixtures
- No limitation on number of contributors
- No limitation on number of replicates that can be combined to form a single LR